



Jonathan Karall  
jaa337@cornell.edu



Neil Mattson  
nsm47@cornell.edu

Volume 9 Number 4 April 2024

## Do Microgreens Respond to Daily Light Integral and Carbon Dioxide Enrichment?

Microgreens are defined as a wide range of vegetable and herb seedlings that are harvested shortly after the emergence of the first true leaf and prior to leaf expansion/senescence of cotyledons. Microgreens represent a quick turn, potentially high value crop for greenhouses and indoor production. Because there are a wide diversity of species used, recommendations for cultural practices are lacking for many. Relatively little information is available on microgreens responses to light and carbon dioxide.



Figure 1. Microgreens have gained popularity as a tasty salad ingredient and garnish.

Because many microgreens start with a relatively large seed that can provide initial nutrition to the developing seedling it is thought they are relatively less responsive to light and CO<sub>2</sub> than their full-size counterparts (ex. baby greens or head lettuce). However, experimental data is needed to evaluate these claims. The objective of this study was to determine how three species of microgreens would respond to daily light integral (DLI) varying from 3 to 12 mol·m<sup>-2</sup>·d<sup>-1</sup> and carbon dioxide enrichment from 400 ppm (ambient) to 1,000 ppm.

### 2024 Sponsors



Research  
Internships  
Scholarships  
Education

Funding the Future of Floriculture

Bali®

fine



P.L. LIGHT SYSTEMS  
THE LIGHTING KNOWLEDGE COMPANY

Reprint with permission from the author(s) of this e-GRO Alert.



Figure 2. Images of microgreen species used in the experiment where A, B and C represent arugula, mizuna and mustard. Images © by Jonathan Karall, Cornell University.

## Experimental methods

Three microgreen species; arugula (*Eruca sativa* L.), mizuna (*Brassica rapa* L. var. japonica) and mustard (*Brassica juncea* ‘Garnet Giant’) were chosen for this study. The seeds were purchased from Johnny’s Seed and were selected to represent a diversity in sensory attributes for both vision and taste. We also choose these three as their share similarities in seed size and number of days from seeding to harvest.

Previous experiments were conducted to determine cultural parameters such as seed density, fertilizer, and substrate ([see GPN article](#)). We used flats with 2401 (24-cell) inserts with a peat-based potting mix (Lambert LM-111) pre watered to a moisture content ratio of 1:1 (peat-lite mix:fertilizer solution) by weight with Jack’s 21-5-20 Liquid Feed at 150 ppm nitrogen. Seeds were sown on the top of the substrate at a rate of 125 seeds per cell (equivalent to 3,000 seeds per 20”x10” flat). Seeds were then misted with the same fertilizer solution. For germination, the trays were covered with a propagation dome that was then covered with a standard black flat to restrict light. Seeds were then germinated in darkness at 73 °F until 95% of the seedlings were 1 cm in height. Germination times were 42 hours for arugula, 46 hours for mizuna, and 48 hours for mustard. After germination, microgreens were placed into the respective treatments (described below) with 12 cells per species per treatment.

The experiment used two adjacent controlled environment chambers at 73 °F, each with their own mini acrylic chambers which allowed for control of CO<sub>2</sub>, temperature, and relative humidity. For each experimental run, one DLI (from 3, 6, 9, and 12 mol·m<sup>-2</sup>·d<sup>-1</sup>) was selected. The growth chambers had T5 cool white fluorescent lights. Light was supplied over a 14 hour photoperiod. Shade cloth (50%) was used over afor the lowest DLI. Each of the four acrylic mini chambers was randomly assigned one of the CO<sub>2</sub> concentrations (400, 600, 800, 1000 ppm). Flats were watered as needed (roughly every 3-days) via subirrigation with Jack’s 21-5-20 at 150 ppm N. The experiment was repeated for each DLI and then replicated over time for a total of three times.



Microgreens were harvested when 50% of seedlings had a first true leaf measuring 1 cm in length. Measured parameters included average plant height (from substrate to the tallest part of representative plants), days to harvest, and fresh weight per cell.

### Fresh weight

For mizuna and mustard there was a linear increase in fresh weight as DLI increased from 3 to 12 mol·m<sup>-2</sup>·d<sup>-1</sup> (Figure 3) Fresh weight increased by 22-25% from the lowest to the highest DLI treatment. Arugula was more responsive to DLI, fresh weight increased by 51% from 3 to 9 mol·m<sup>-2</sup>·d<sup>-1</sup> and then exhibited little further gains from 9 to 12 mol·m<sup>-2</sup>·d<sup>-1</sup>. All species responded to carbon dioxide enrichment in a linear fashion showing about an 11% yield increase from 400 to 1,000 ppm CO<sub>2</sub>. Put another way, CO<sub>2</sub> enrichment could be used to reduce the amount of lighting needed. For example, fresh weight of mizuna at a DLI of 9 mol·m<sup>-2</sup>·d<sup>-1</sup> with 1,000 ppm CO<sub>2</sub> was greater than a DLI of 12 mol·m<sup>-2</sup>·d<sup>-1</sup> with ambient CO<sub>2</sub> (400 ppm).

### Height

Average plant height for mizuna and mustard decreased linearly as DLI increased from 3 to 12 mol·m<sup>-2</sup>·d<sup>-1</sup> (Figure 4). Decreases amounted to reductions in plant height of 0.9 cm for mizuna and 1.3 cm for mustard as DLI increased from 3 to 12 mol·m<sup>-2</sup>·d<sup>-1</sup> at 400 ppm CO<sub>2</sub>.

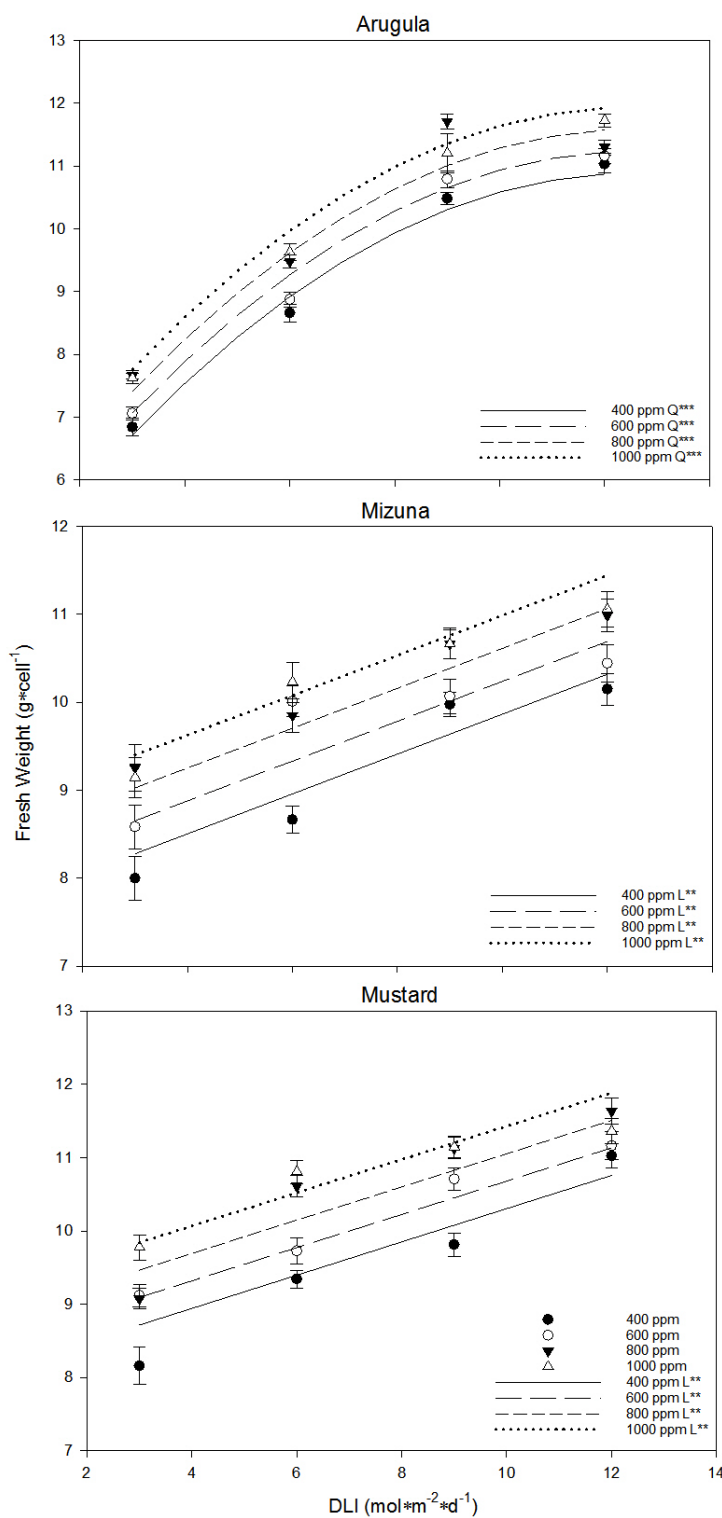


Figure 3. Fresh weight (grams per cell) of arugula, mizuna, and mustard microgreens in response to daily light integral and CO<sub>2</sub> (Note 1 20"x10" tray = 24 cells)

For arugula, plant height was essentially unaffected by DLI but did decrease at the highest treatment 9 mol·m<sup>-2</sup>·d<sup>-1</sup>. CO<sub>2</sub> also had a subtle effect where plant height increased slightly (0.4 cm averaged across species and light levels) as CO<sub>2</sub> increased from 400 to 1000 ppm.

### Days to Harvest

The days to harvest varied by species and was about 12 days for arugula and mustard and 10.5 days for mizuna at the lowest DLI (Figure 5). Days to harvest decreased for all species by about two days as DLI increased to 12 mol·m<sup>-2</sup>·d<sup>-1</sup>. The CO<sub>2</sub> concentration did not influence plant developmental stage (i.e. days from seeding to harvest).

### Bottom Line

While microgreens do not appear to require DLIs as high as mature leafy greens crops (ex. 17 mol·m<sup>-2</sup>·d<sup>-1</sup> for head lettuce) or fruiting crops (ex. 25+ mol·m<sup>-2</sup>·d<sup>-1</sup> for tomatoes), microgreens may benefit from increasing DLI up to 9 mol·m<sup>-2</sup>·d<sup>-1</sup> for arugula and 12 mol·m<sup>-2</sup>·d<sup>-1</sup> (and potentially beyond) for mizuna and mustard. Growers should understand their ambient DLI (such as by purchasing a quantum sensor connected to a datalogger) to then assess if supplemental light makes sense (or during which months it makes sense). Besides increasing fresh weight, increased DLI reduced days to harvest by about two days resulting in quicker crop turns. CO<sub>2</sub> enrichment from ambient to 1,000 ppm increased harvested fresh weight by 11%. Thus, growers should

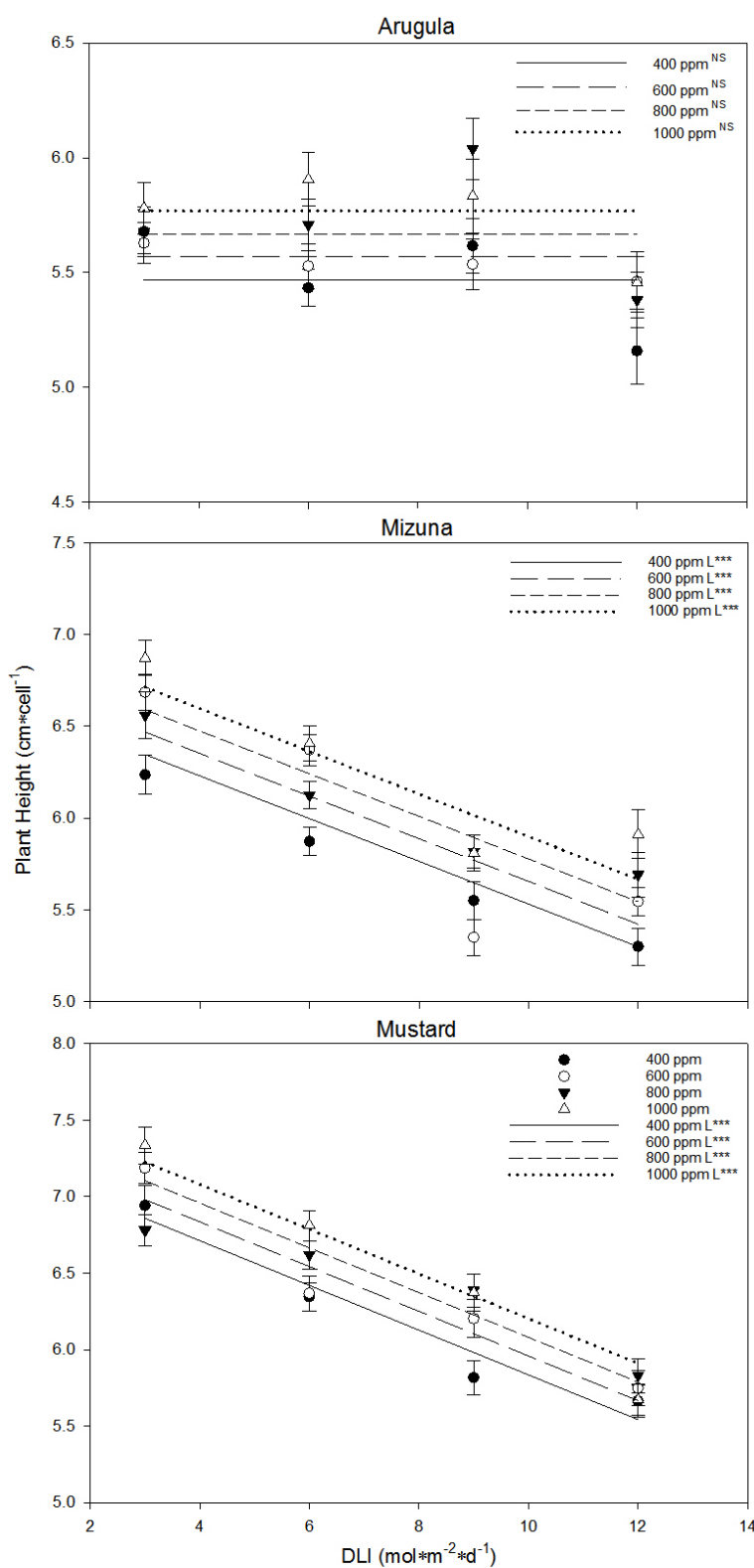


Figure 4. Height of arugula, mizuna, and mustard microgreens in response to daily light integral and CO<sub>2</sub>.

assess the economics of CO<sub>2</sub> enrichment and in closed production systems (i.e. no open ventilation) there may be an economic returning of CO<sub>2</sub> enrichment. Plant height decreased by up to 1.3 cm with increasing DLI. For some growers, if microgreens are too compact in height they can be difficult to harvest. If these growers use higher light intensities they may want to consider measures to increase stem height such as a slightly longer germination period in the darkness. More research is needed to quantify how other microgreens species/cultivars respond to light and carbon dioxide. Commercial growers should always conduct small-scale trials to see how plants respond in their own facilities before adopting new practices.

Overall microgreens can be a profitable crop and they may be more practical (in terms of energy cost) for indoor farming without sunlight as they require lower DLI than head lettuce and fruiting crops.

**Citations**

Allred, J. and N. Mattson. 2018. Growing better greenhouse microgreens. Greenhouse Product News, Under Control, CEA supplement. October:10-13. [Available online.](#)

Allred, J. 2017. Environmental and cultural practices to optimize the growth and development of three microgreen species. M.S. Thesis. Cornell University. 78pp. [Available online.](#)

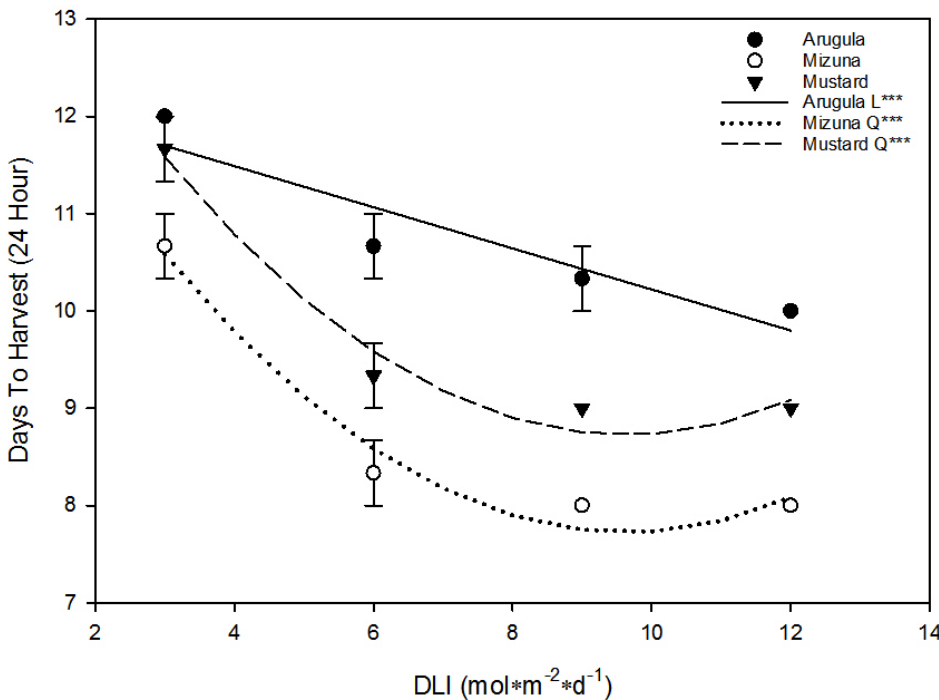


Figure 5. Days from seed to harvest of arugula, mizuna, and mustard microgreens in response to daily light integral and CO<sub>2</sub>.



**e-GRO Alert**

[www.e-gro.org](http://www.e-gro.org)

**CONTRIBUTORS**

Dr. Nora Catlin  
Floriculture Specialist  
Cornell Cooperative Extension  
Suffolk County  
[nora.catlin@cornell.edu](mailto:nora.catlin@cornell.edu)

Dr. Chris Currey  
Assistant Professor of Floriculture  
Iowa State University  
[ccurrey@iastate.edu](mailto:ccurrey@iastate.edu)

Dr. Ryan Dickson  
Greenhouse Horticulture and  
Controlled-Environment Agriculture  
University of Arkansas  
[ryand@uark.edu](mailto:ryand@uark.edu)

Dan Gilrein  
Entomology Specialist  
Cornell Cooperative Extension  
Suffolk County  
[dog1@cornell.edu](mailto:dog1@cornell.edu)

Dr. Chieri Kubota  
Controlled Environments Agriculture  
The Ohio State University  
[kubota.10@osu.edu](mailto:kubota.10@osu.edu)

Heidi Lindberg  
Floriculture Extension Educator  
Michigan State University  
[wolleage@anr.msu.edu](mailto:wolleage@anr.msu.edu)

Dr. Roberto Lopez  
Floriculture Extension & Research  
Michigan State University  
[rglopez@msu.edu](mailto:rglopez@msu.edu)

Dr. Neil Mattson  
Greenhouse Research & Extension  
Cornell University  
[neil.mattson@cornell.edu](mailto:neil.mattson@cornell.edu)

Dr. W. Garrett Owen  
Sustainable Greenhouse & Nursery  
Systems Extension & Research  
The Ohio State University  
[owen.367@osu.edu](mailto:owen.367@osu.edu)

Dr. Rosa E. Raudales  
Greenhouse Extension Specialist  
University of Connecticut  
[rosa.raudales@uconn.edu](mailto:rosa.raudales@uconn.edu)

Dr. Alicia Rihn  
Agricultural & Resource Economics  
University of Tennessee-Knoxville  
[arihn@utk.edu](mailto:arihn@utk.edu)

Dr. Debalina Saha  
Horticulture Weed Science  
Michigan State University  
[sahadeb2@msu.edu](mailto:sahadeb2@msu.edu)

Dr. Beth Scheckelhoff  
Extension Educator - Greenhouse Systems  
The Ohio State University  
[scheckelhoff.11@osu.edu](mailto:scheckelhoff.11@osu.edu)

Dr. Ariana Torres-Bravo  
Horticulture / Ag. Economics  
Purdue University  
[torres2@purdue.edu](mailto:torres2@purdue.edu)

Dr. Brian Whipker  
Floriculture Extension & Research  
NC State University  
[bwhipker@ncsu.edu](mailto:bwhipker@ncsu.edu)

Dr. Jean Williams-Woodward  
Ornamental Extension Plant Pathologist  
University of Georgia  
[jwoodwar@uga.edu](mailto:jwoodwar@uga.edu)

Copyright ©2024

Where trade names, proprietary products, or specific equipment are listed, no discrimination is intended and no endorsement, guarantee or warranty is implied by the authors, universities or associations.

**Cooperating Universities**

**Cornell CALS**  
College of Agriculture and Life Sciences

**Cornell Cooperative Extension  
Suffolk County**

**UTIA INSTITUTE OF  
AGRICULTURE**  
THE UNIVERSITY OF TENNESSEE

**IOWA STATE UNIVERSITY**



**College of Agricultural &  
Environmental Sciences**  
UNIVERSITY OF GEORGIA

**UCONN**

**UofA DIVISION OF AGRICULTURE  
RESEARCH & EXTENSION**  
University of Arkansas System

**MICHIGAN STATE  
UNIVERSITY**

**NC STATE  
UNIVERSITY**

**P PURDUE  
UNIVERSITY**



**THE OHIO STATE  
UNIVERSITY**

**In cooperation with our local and state greenhouse organizations**

**MAUMEE VALLEY GROWERS**  
Choose the Very Best.



**Metro Detroit Flower Growers Association**

